

# De la chimie à la biologie: les nouvelles frontières de l'analyse des médicaments de dernière génération

26<sup>ème</sup> Journée Scientifique du cCCTA « Biologie-Chimie, l'entre deux mondes... »  
14 – 15 septembre, Villars-sur-Ollon, Suisse

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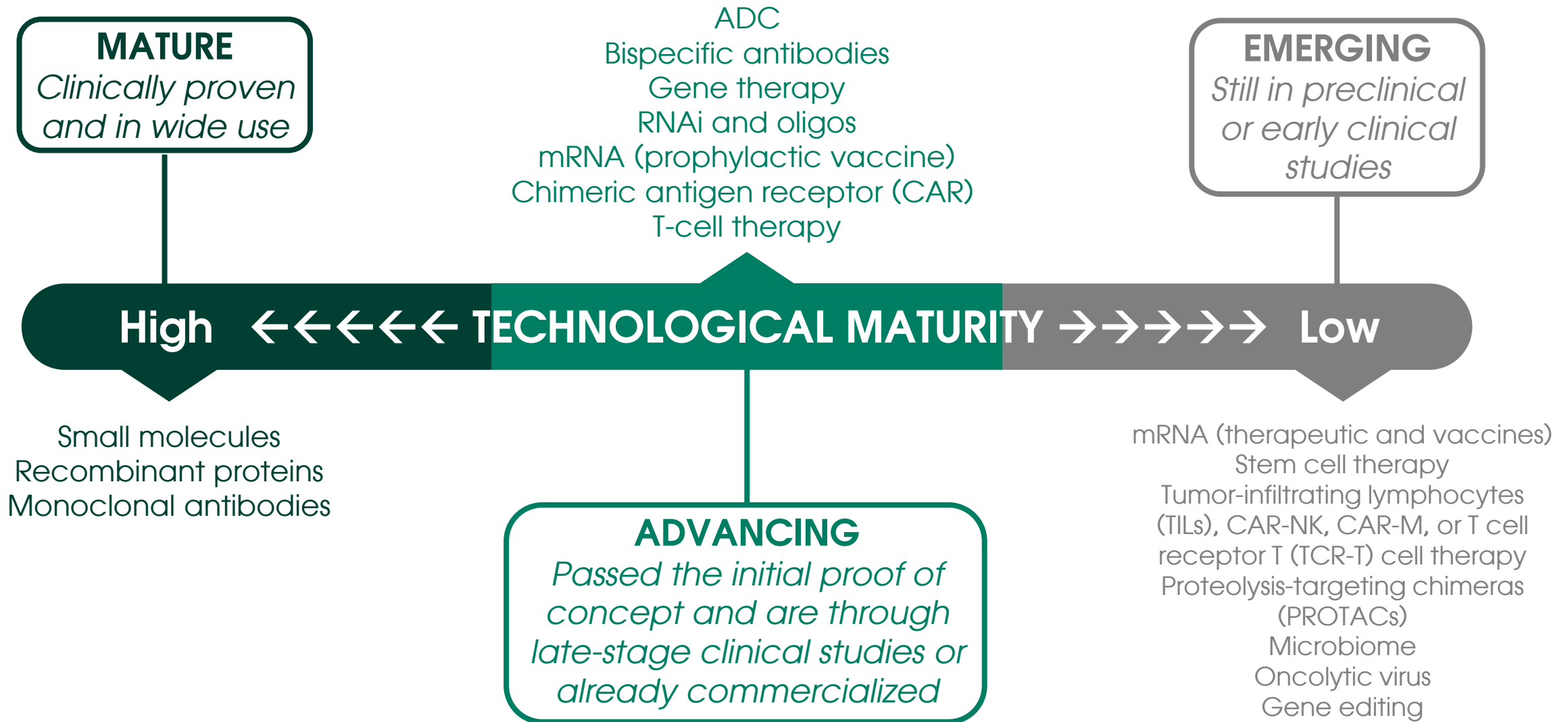
FACULTÉ DES SCIENCES  
SECTION DES SCIENCES PHARMACEUTIQUES

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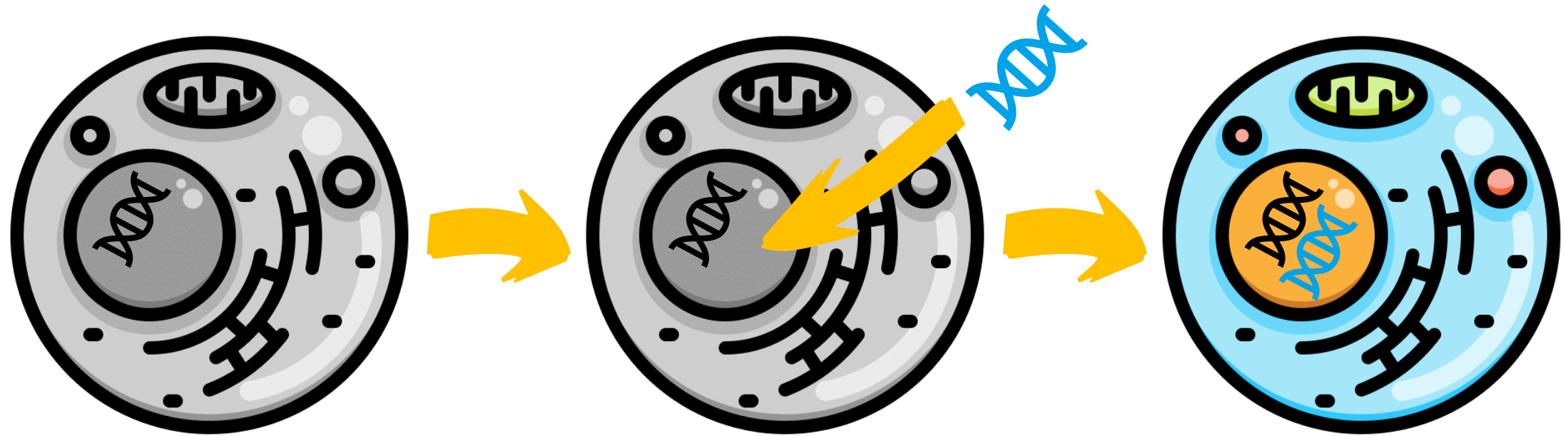
# Where are we in terms of new modalities?



<https://www.bcg.com/publications/2023/benefits-and-risks-of-new-drug-modalities>

# What is gene therapy?

Therapy that targets the genetic root cause of a specific disease



Cell has defective gene

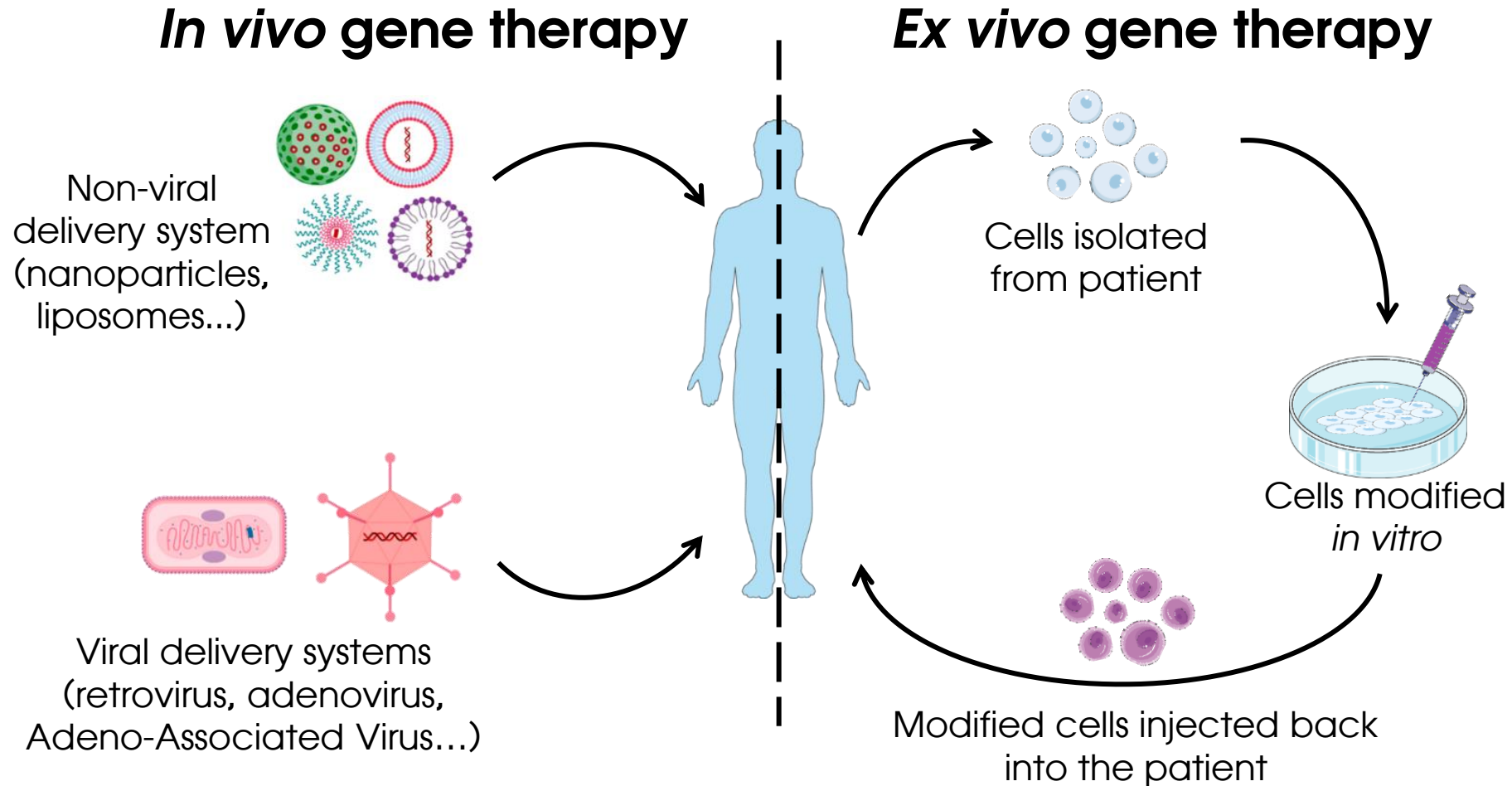
Healthy gene is introduced

Cell function is restored

A working copy of the defective gene is delivered as therapeutic drug

# How to deliver a therapeutic gene?

Two different strategies are possible for delivering a therapeutic gene



# Timeline of the FDA-approved gene therapies

Three AAV-based therapies currently approved by the FDA

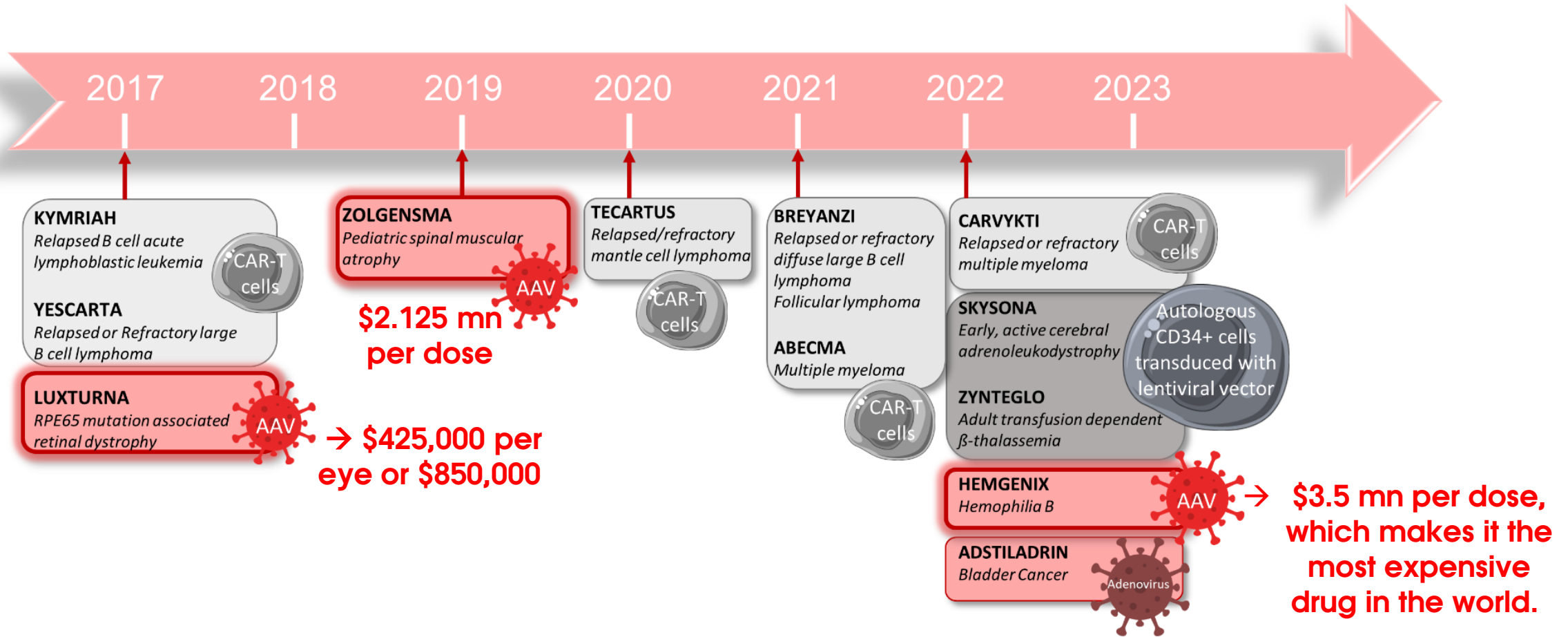


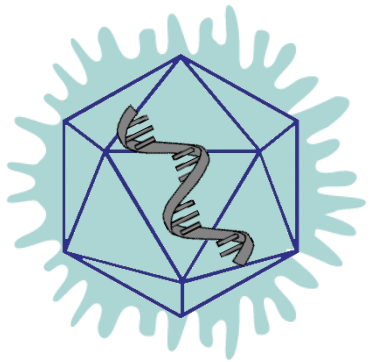
Figure ref = Fekete et al. TrAC 2023, 117088

\* <https://www.fda.gov/vaccines-blood-biologics/cellular-gene-therapy-products/approved-cellular-and-gene-therapy-products>



# Adeno-Associated Virus (AAV) as vectors for gene delivery

AAV are a safe and attractive vectors for gene delivery



Non-pathogenic single stranded DNA virus



Small viral genome (ss DNA < 4.8 kb)



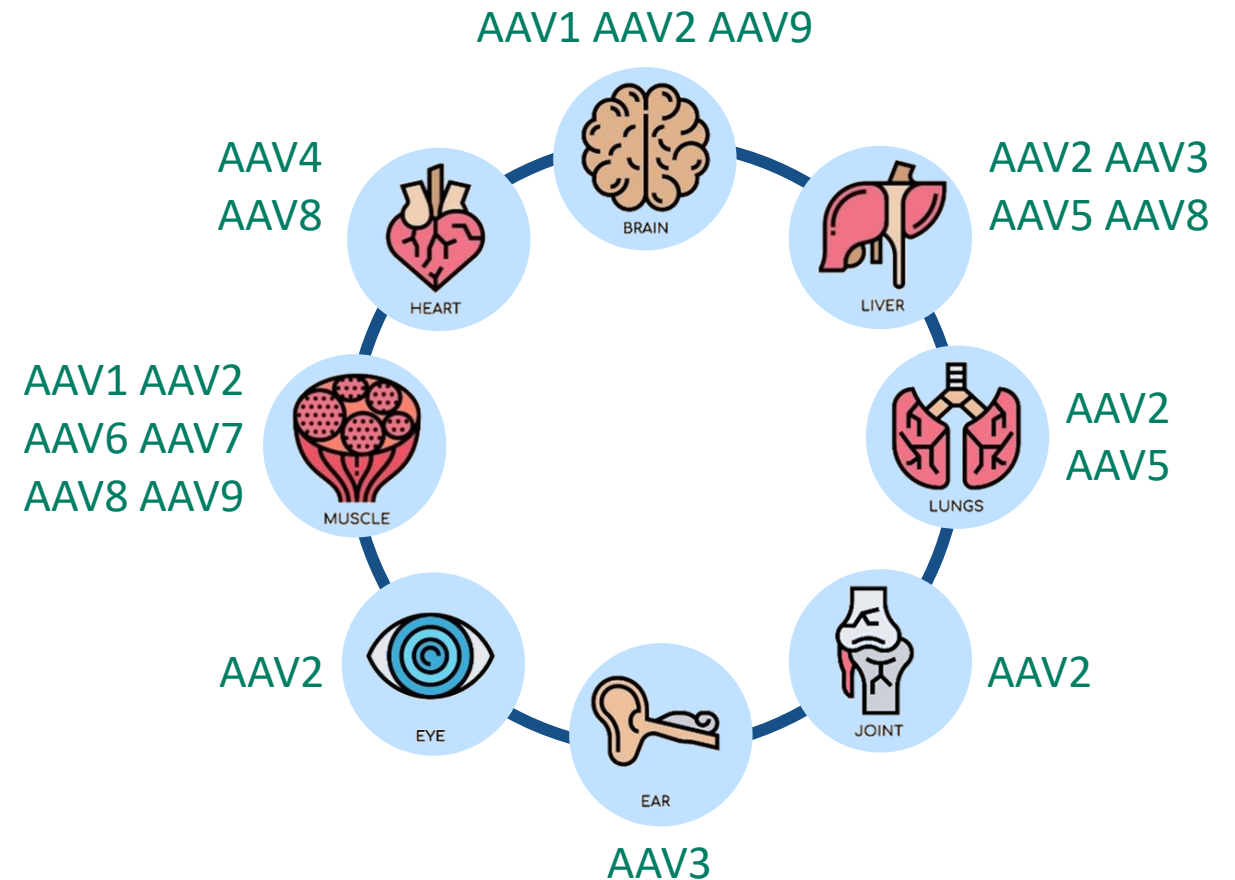
Icosahedric capsid consisting of ~60 viral proteins (VP1, VP2, VP3)



Diameter = 22 nm  
Mass = 3700 kDa



12 different serotypes



AAVs serotype have specific target tissue tropism

# Analytical methods to be established for AAV-based gene therapy

Parameter	Method
Full/Empty Capsid	AUC (gold standard), TEM (gold standard), IEX, SEC-MALS, CE-SDS/cIEF
Aggregation and Fragmentation	SEC, SEC-MALS, DLS
Capsid Integrity (VP1-3 Ratio)	RPLC, CE-SDS/cIEF
Capsid Identity (VP1-3 Ratio), PTM	LC-MS
Particle Concentration (viral titer)	SEC-MALS, ELISA
Genomic titer, genomic identity	qPCR/ddPCR
Infectivity, potency, mode of action	Cell based assays

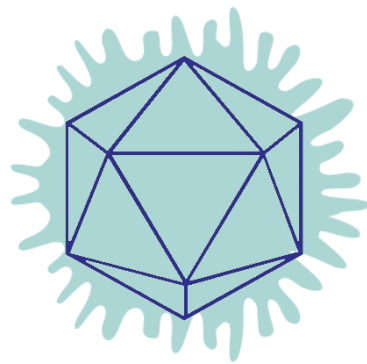
AUC = Analytical Ultra Centrifugation; TEM = Transmission Electron Microscopy; IEX = Ion Exchange Chromatography; SEC = Size Exclusion Chromatography; MALS = Multi-angle Light Scattering; DLS = Dynamic Light Scattering; ELISA = Enzyme-Linked Immunosorbent Assay; RPLC = Reversed-Phase Liquid Chromatography; CE-SDS = Capillary electrophoresis sodium dodecyl sulfate; cIEF = capillary IsoElectric Focusing; q/ddPCR = quantitative/droplet digital Polymerase Chain Reaction

# IEX method to quantify Full vs Empty capsids

Empty rAAV capsids represent a product-related impurity that need to be monitored and quantified as represent a safety concern.

Generic salt-mediated AEX gradient conditions for rAAV Empty and Full capsids separation

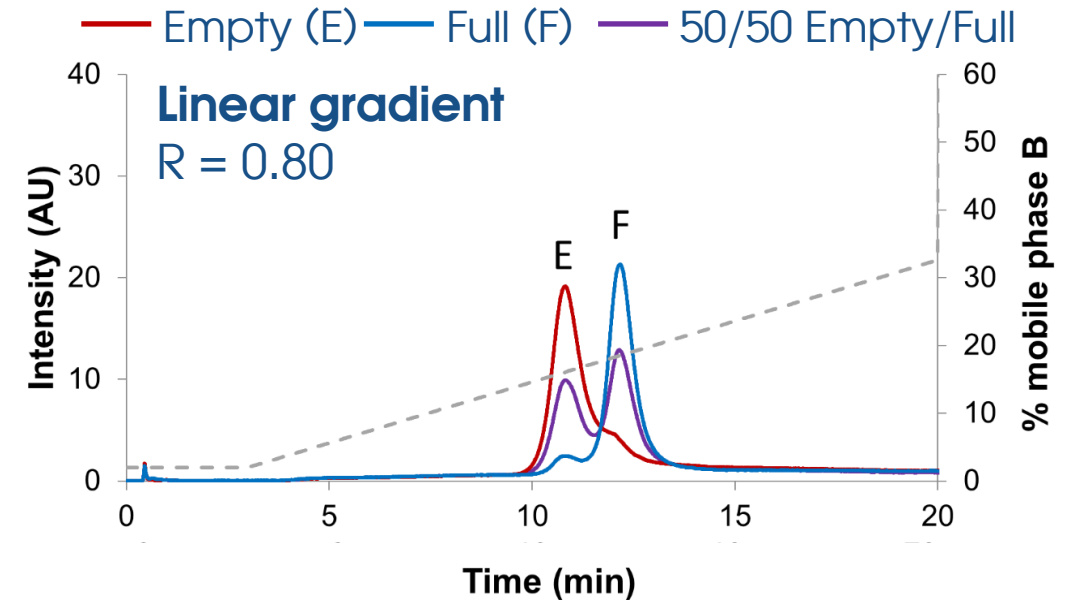
## Product related impurity



EMPTY

- Reduce effective concentration
- Compete for binding sites
- Increase immunogenicity
- Reduce efficacy

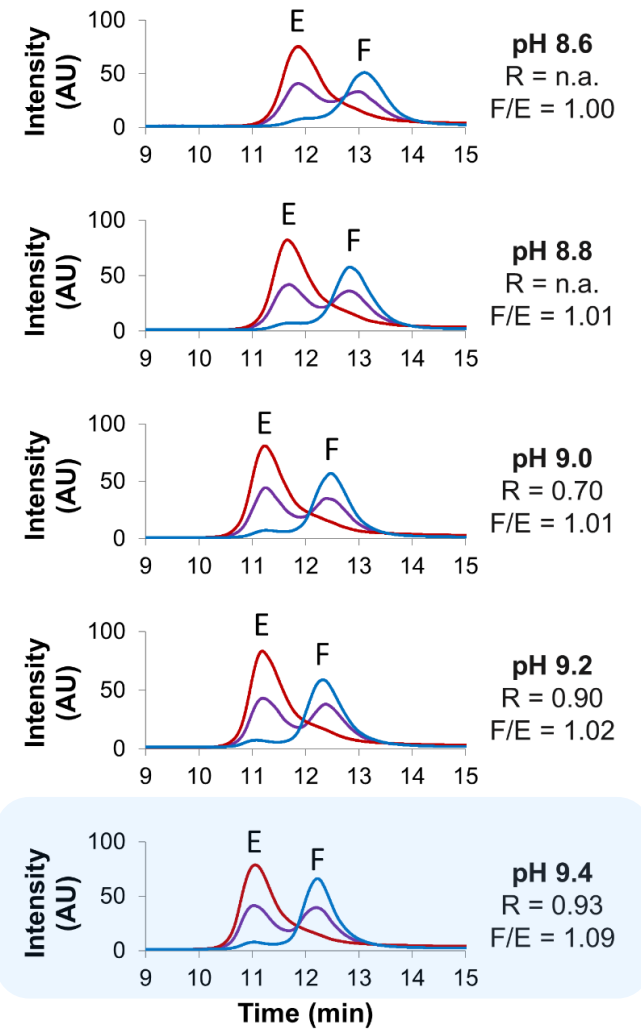
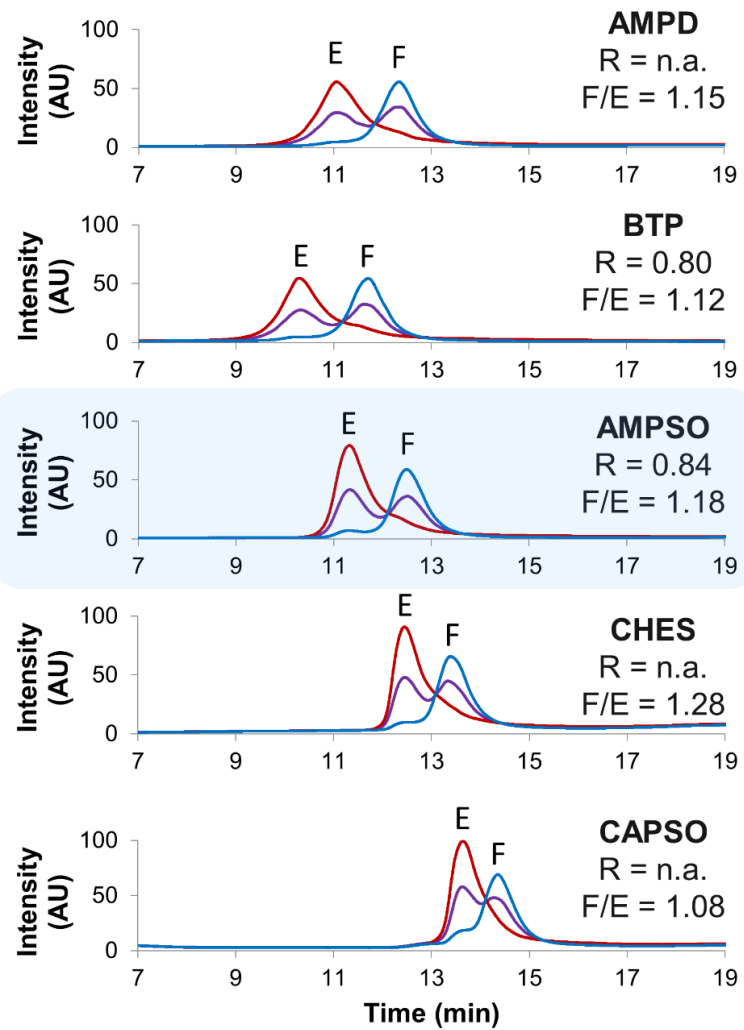
Issue = Empty and Full rAAV capsids have the same size and minor pI difference (0.4 pH) so poor chromatographic resolution is generally obtained.



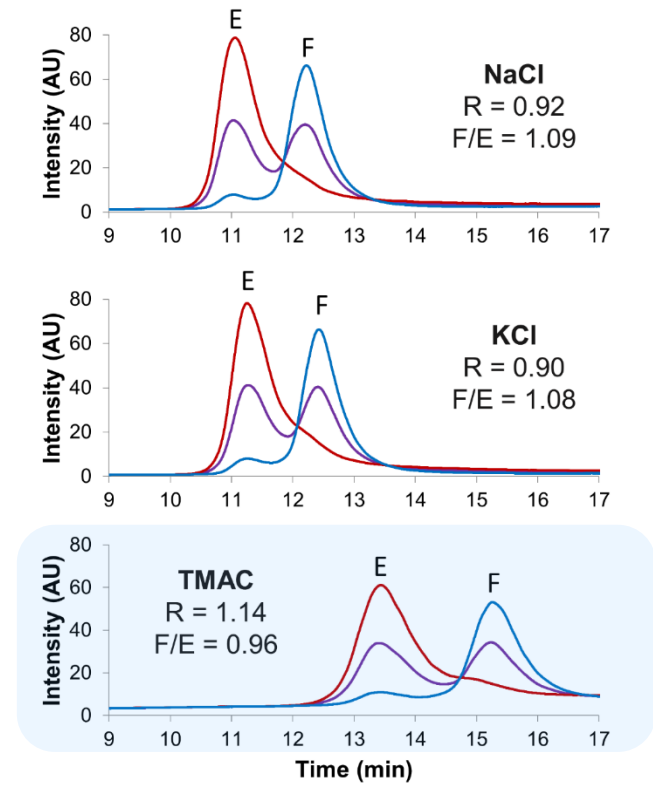
Gradient conditions = linear gradient of BTP buffer pH 9.0 in combination with NaCl as eluent salt. FLD detection =  $\lambda_{ex}$  at 280 nm and  $\lambda_{em}$  at 350 nm.  
Injection volume = 20  $\mu$ L of 1.00 E+12 vp/mL samples diluted in water.  
Column = Thermo ProPac SAX-10 50 x 4 mm, 10  $\mu$ m (nonporous PEEK column hardware)



# Buffer scouting plus pH and salt screening



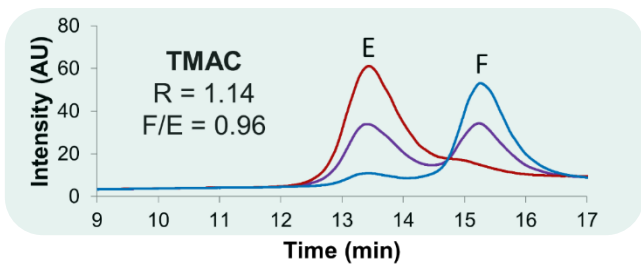
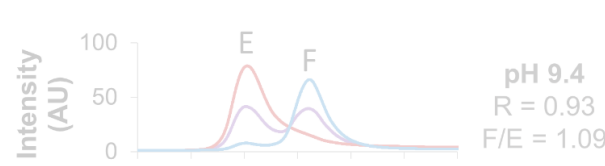
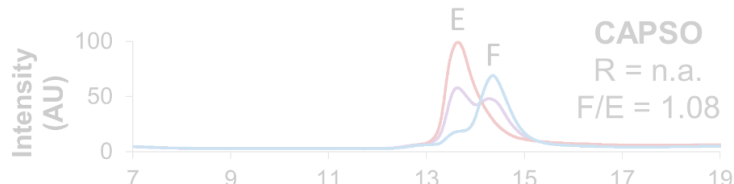
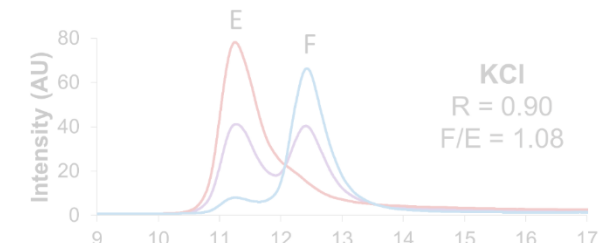
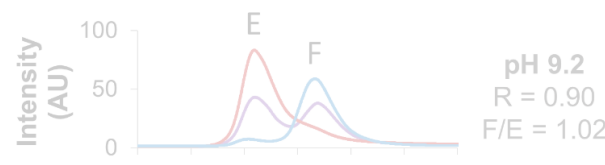
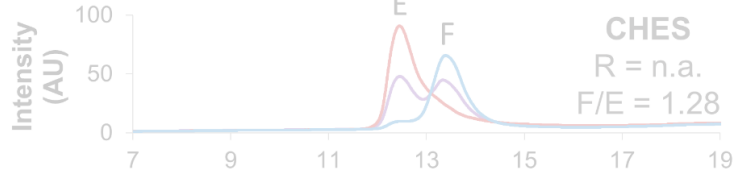
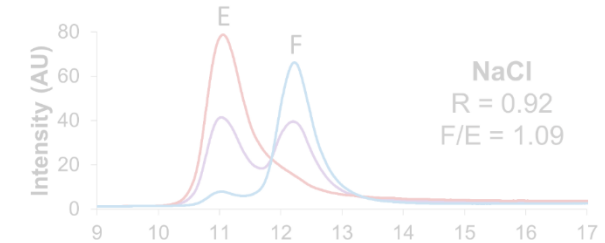
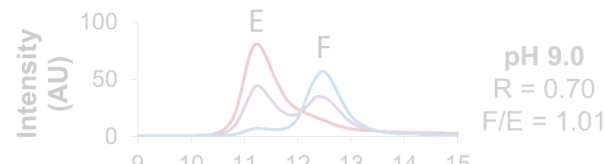
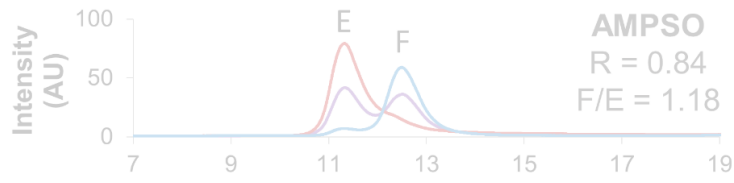
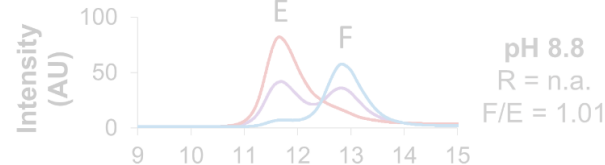
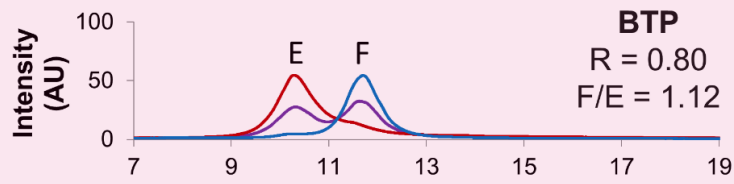
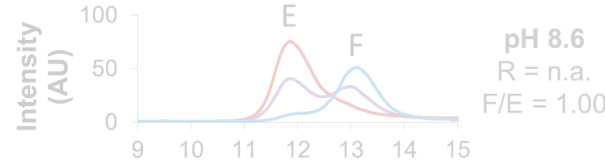
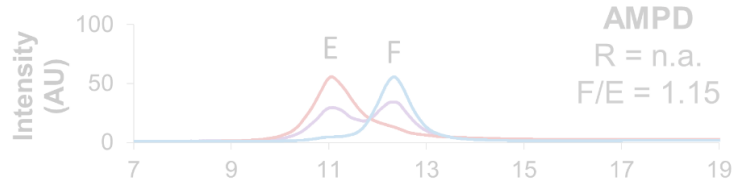
Higher resolution obtained when using tetramethylammonium chloride (TMAC) as eluent salt.



→ Gradient conditions: linear gradient of AMPSO buffer pH 9.4 in combination with different eluent salts.

— Empty (E) — Full (F) — 50/50 Empty/Full

# Linear gradient optimization is not enough



→ Gradient conditions: linear gradient of AMPSO buffer pH 9.4 in combination with different eluent salts.

— Empty (E) — Full (F) — 50/50 Empty/Full

## AAV capsids are made of proteins, do they follow an "on/off" retention behavior?

LSS model describes the relationship between solute retention ( $k$ ) and mobile phase composition ( $\varphi$ )

$$\log k = \log k_0 - S \times \varphi$$

$k$  = retention factor

$\varphi$  = volume fraction of mobile phase "B"  
(stronger eluent)

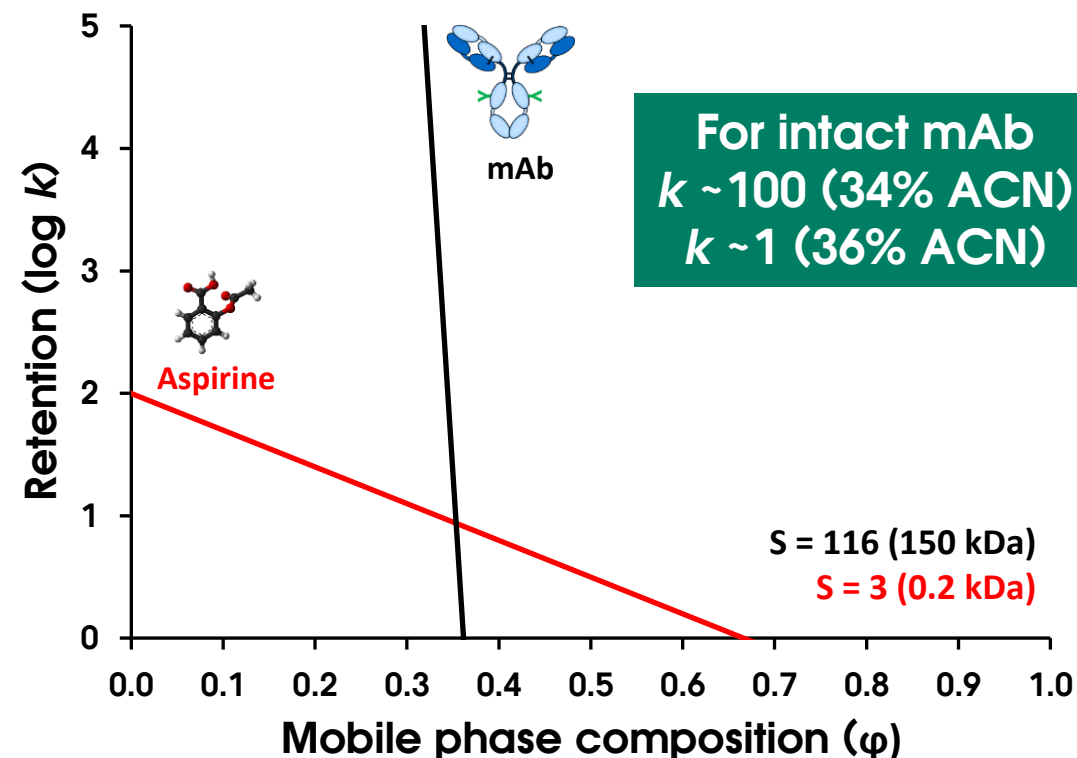
$S$  = constant for a given solute  
(describes how sensitive is the retention to  $\varphi$ )

$k_0$  = (extrapolated) value of  $k$  for  $\varphi = 0$

The slope of these curves drastically increases with the size of the molecule

Large molecules follow an "ON-OFF" or "Bind-and-elute" retention mechanism

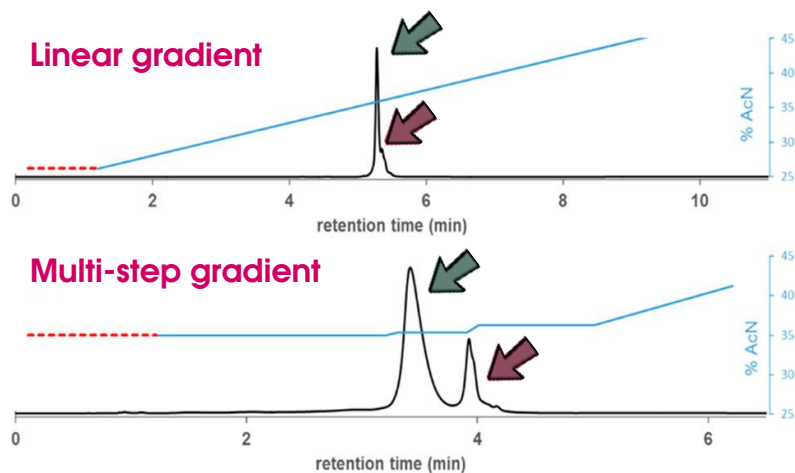
### Linear Solvent Strength (LSS)



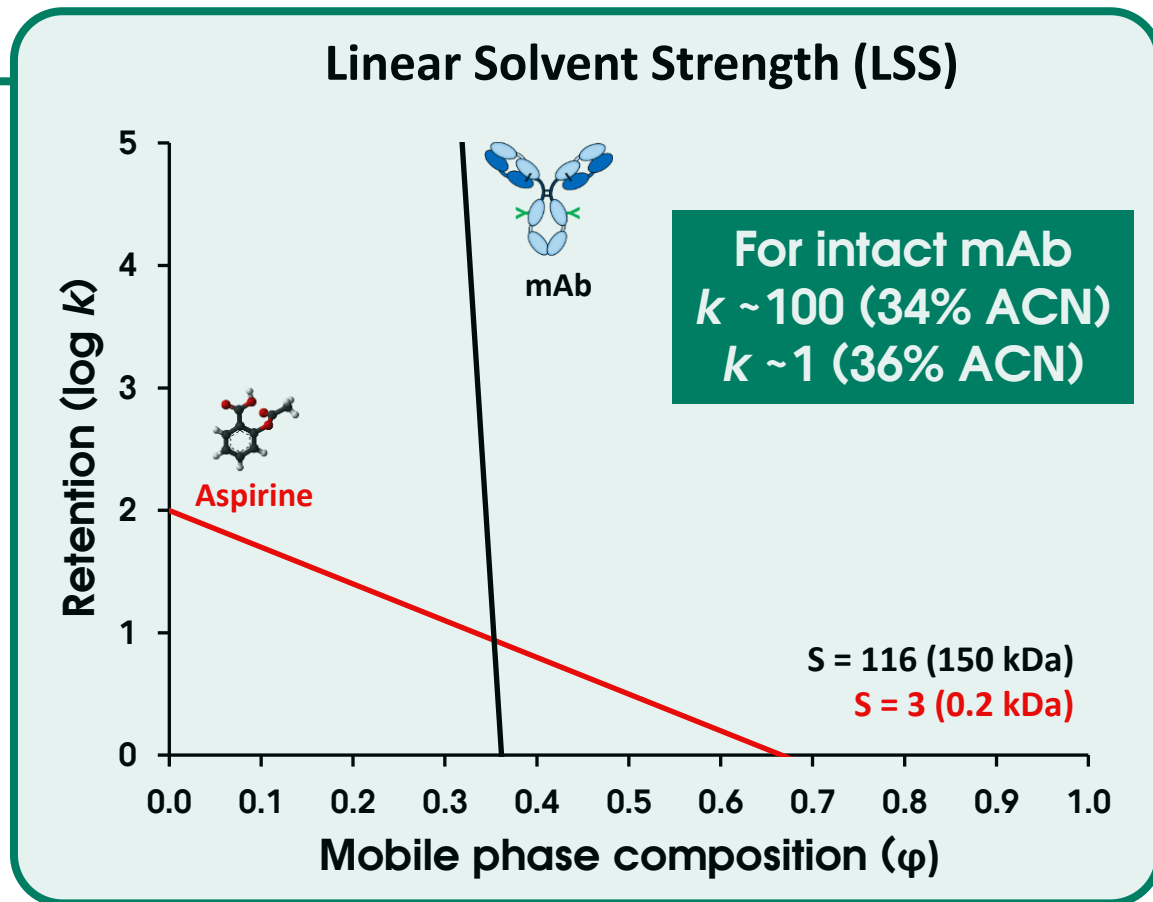
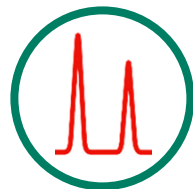
## AAV capsids are made of proteins, do they follow an “on/off” retention behavior?

Large molecules (proteins) are not eluted until exposed to a given mobile phase composition!

Selectivity can be arbitrarily changed between peak pairs by alternating several gradients and isocratic steps.



Fekete et al. *Analytical Chemistry* 2021, 93, 1277-1284

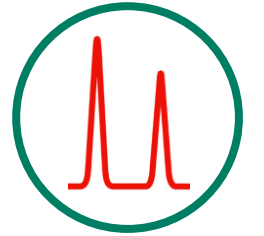


Fekete et al. *Analytical Chemistry* 2019, 91, 12954-12961

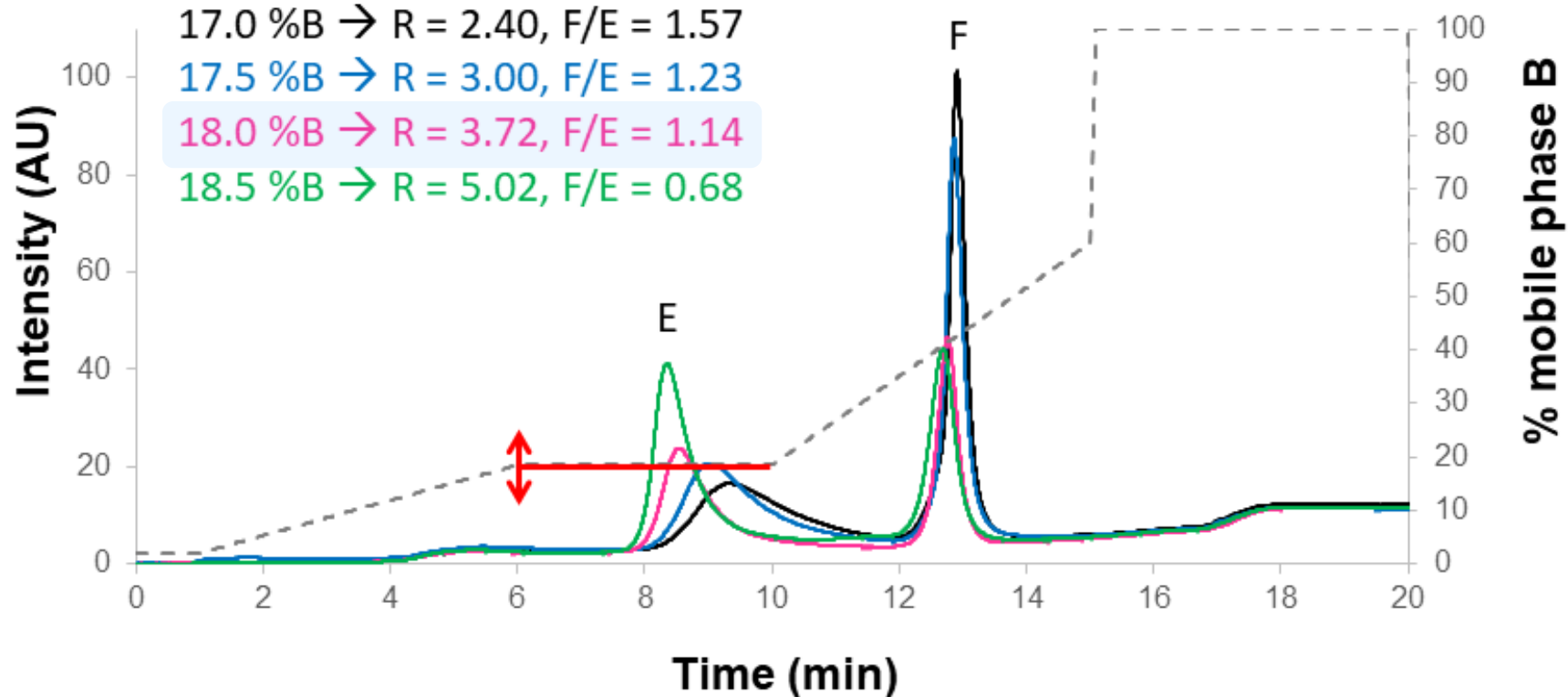
# Improved selectivity by using multi-isocratic elution



AAV capsids are made of proteins,  
do they follow an “on/off” retention behavior?



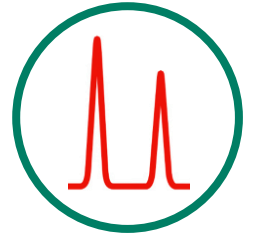
Optimal gradient conditions modeled by DryLab → Isocratic step at 17.5 %B.



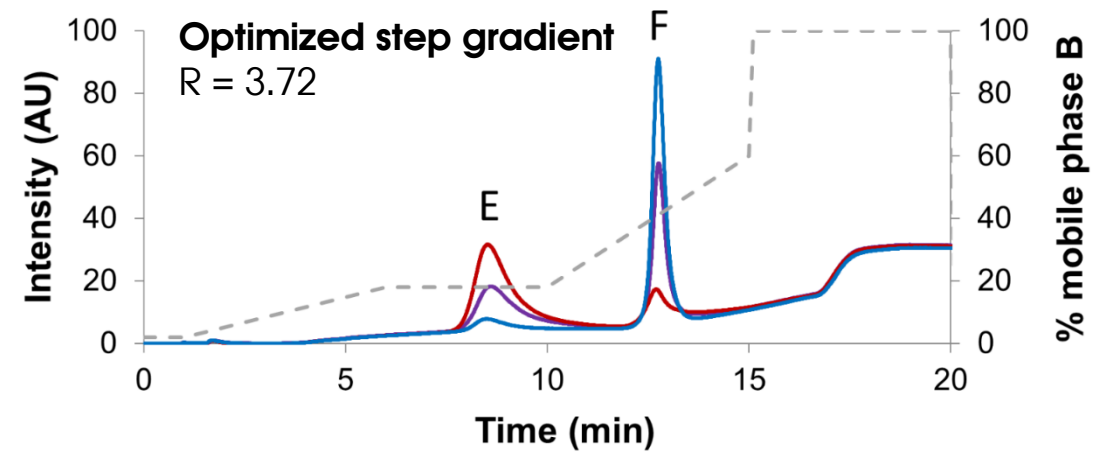
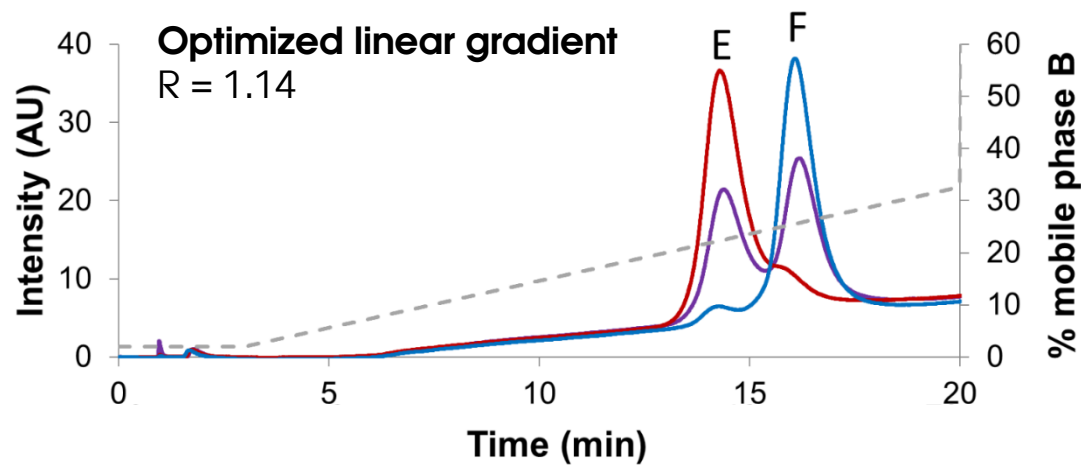
# Improved selectivity by using multi-isocratic elution



AAV capsids are made of proteins,  
do they follow an “on/off” retention behavior?



Optimal gradient conditions modeled by DryLab → Isocratic step at 17.5 %B.



Remarkable improved resolution obtained when  
applying the step gradient method!

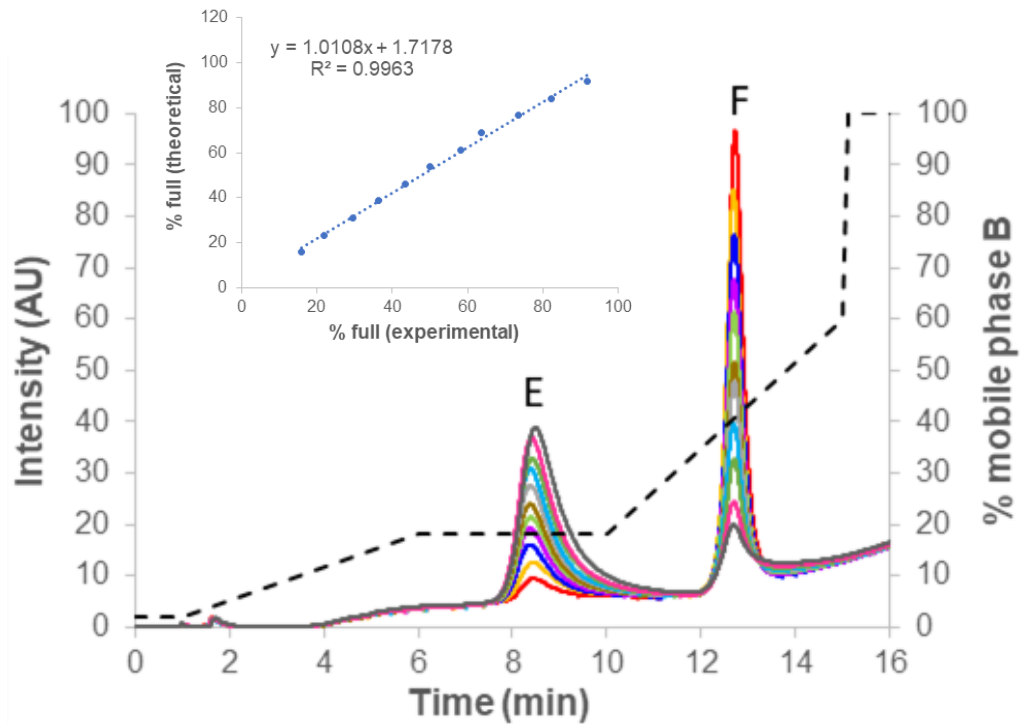
— Empty (E) — Full (F) — 50/50 Empty/Full



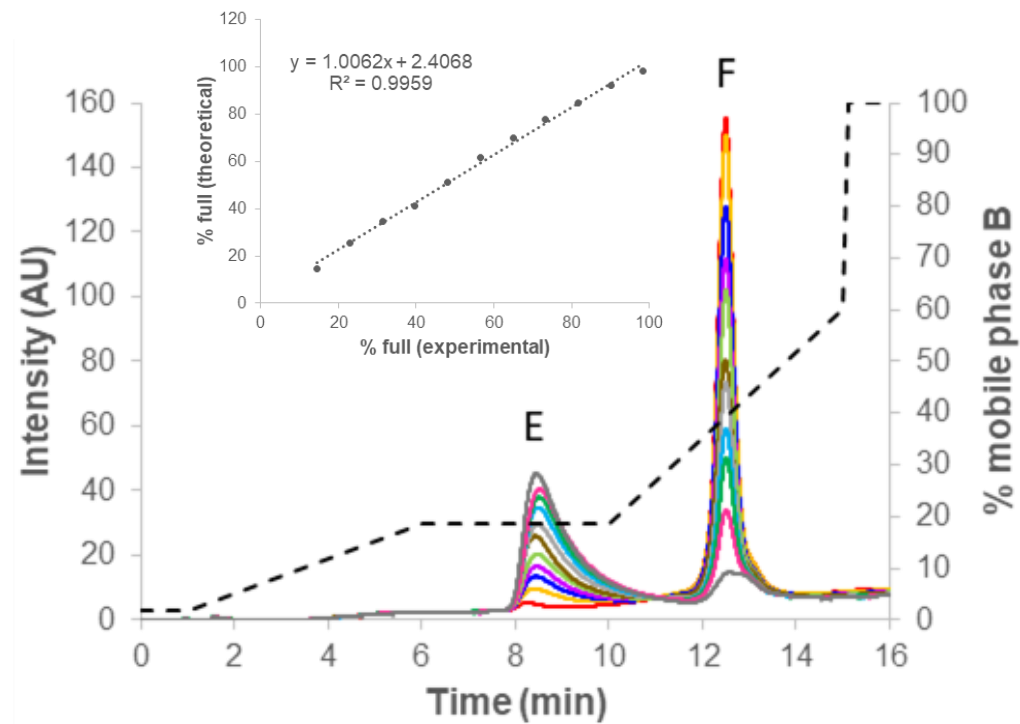
# Method validation by Full/Empty quantification



## Samples from Virovek



## Samples from Sirion Biotech

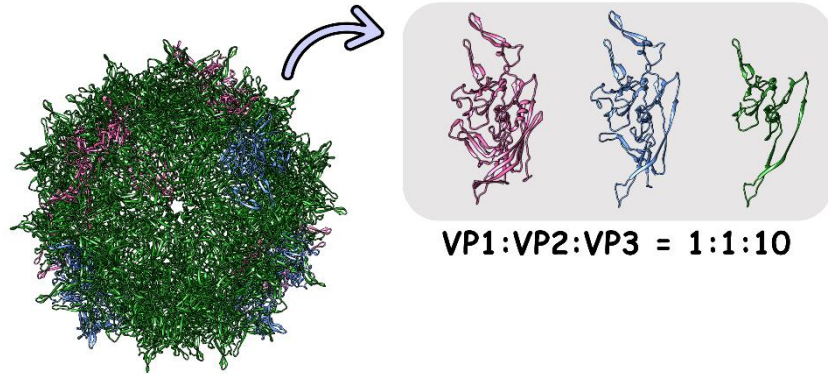


- E/F 0:100
- E/F 10:90
- E/F 20:80
- E/F 30:70
- E/F 40:60
- E/F 50:50
- E/F 60:40
- E/F 70:30
- E/F 80:20
- E/F 90:10
- E/F 100:0
- - - %B

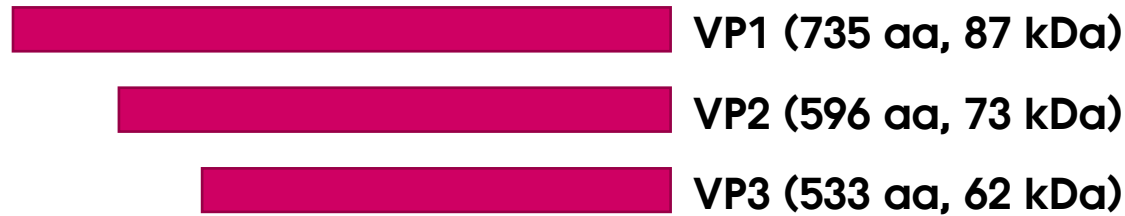
Evaluation of the method from a quantitative perspective:  
the area of each peak was additive, linear and specific to the empty and full rAAV

**The developed method has the potential to be used in QC environment!!**

# Capsid integrity evaluation (VP1-3 Ratio)

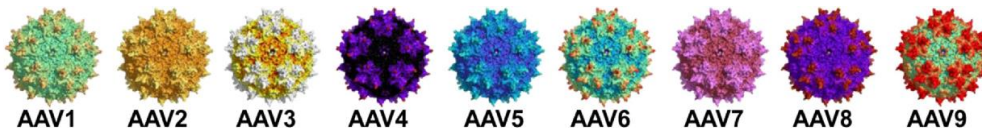


Intact AAV

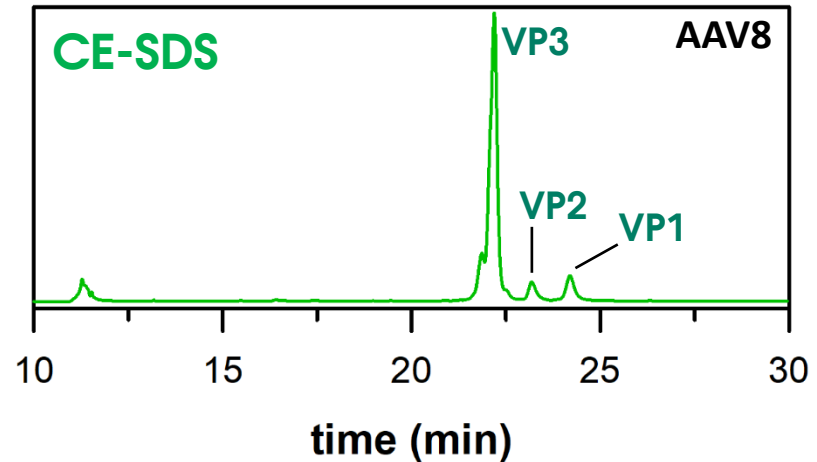


Common PTMs = phosphorylation, deamidation, oxidation, acetylation...

Large sequence homology (51 – 99%) among VPs derived from different AAV serotypes



VPs are generally analyzed by CE-based techniques.  
CE-SDS/cIEF are routinely applied for their excellent resolving power of the VPs.

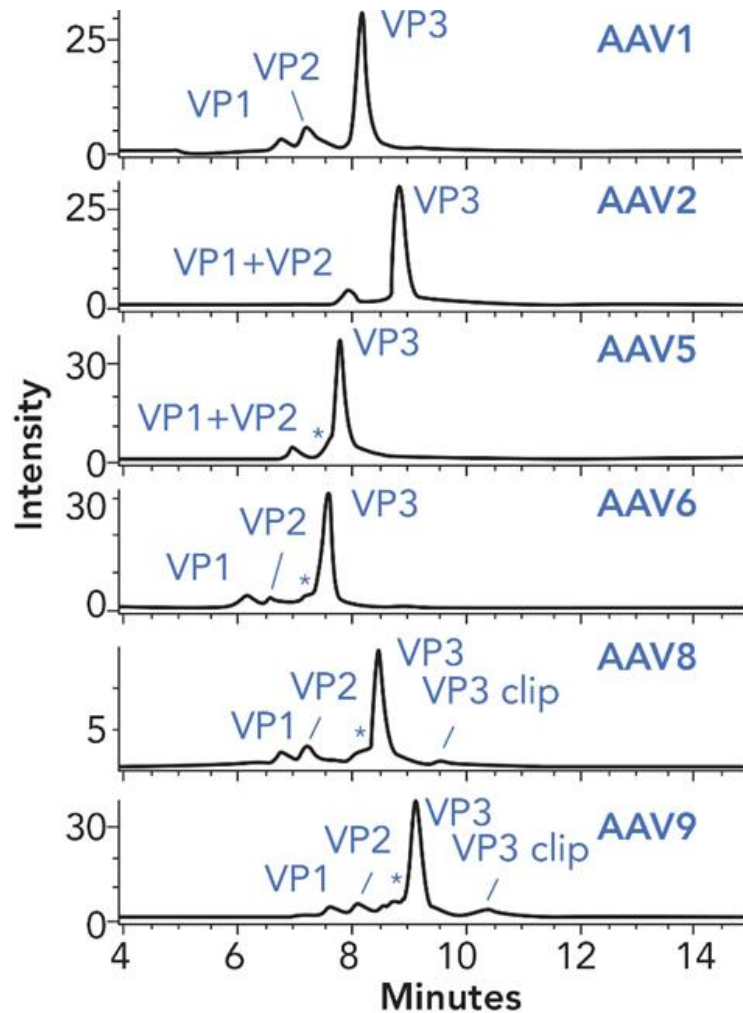


CE-SDS = Capillary electrophoresis sodium dodecyl sulfate; cIEF = capillary IsoElectric Focusing

# Capsid integrity evaluation by LC-based methods

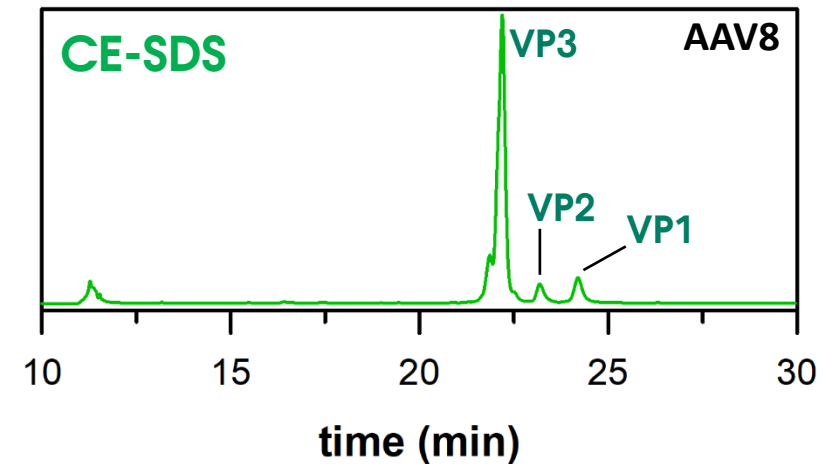


## RPLC



RPLC mode Waters Protein BEH C4 column (300A, 1,7  $\mu$ m, 2,1x150mm); Gradient 32% to 36%B in 16 min. MPA = 0,1% DFA in water, MPB = 0,1% DFA in ACN. FR = 0,2 mL/min, T = 80°C.

VPs are generally analyzed by CE-based techniques.  
CE-SDS/cIEF are routinely applied for their excellent resolving power of the VPs.



← RPLC/MS is used as valid orthogonal approach!

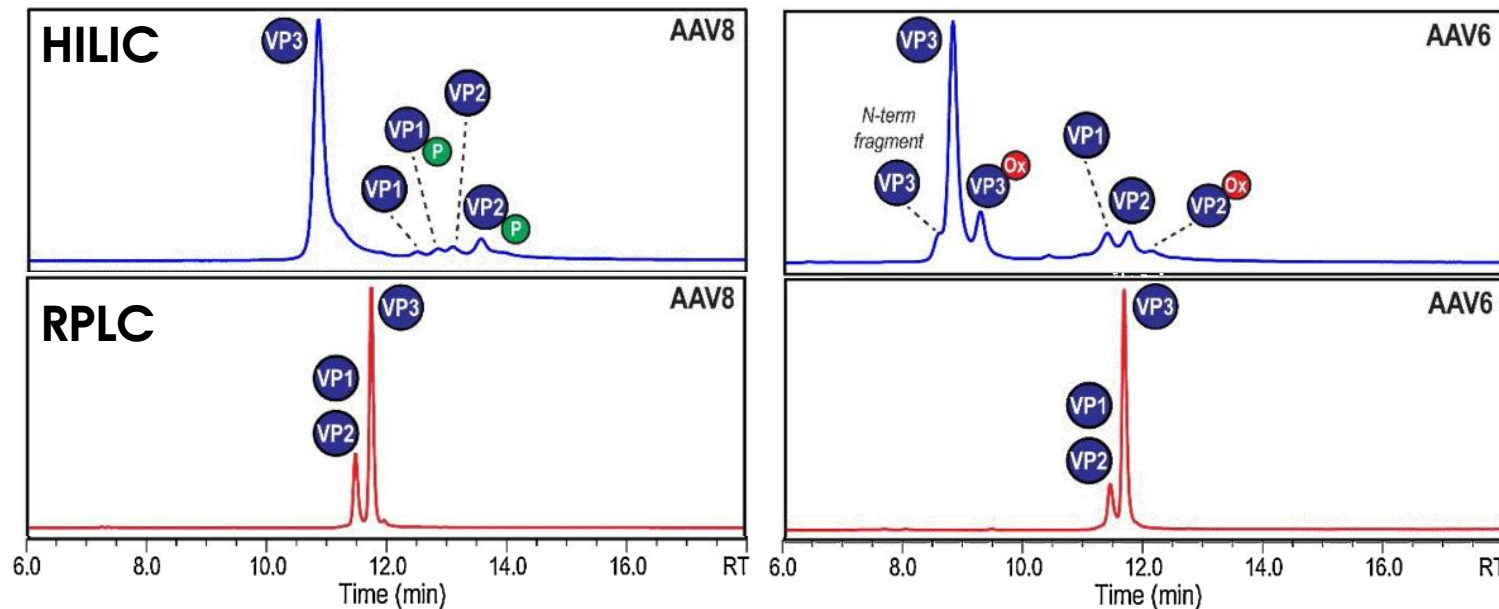
REF RPLC = Zhang et al. Human Gene Therapy 2021, 32, 1501-1511;

CE-SDS = Capillary electrophoresis sodium dodecyl sulfate; cIEF = capillary IsoElectric Focusing

# Improved selectivity by using multi-isocratic elution

HILIC is an extremely interesting option as it has a complementary retention mechanism to RPLC.

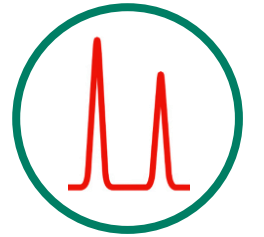
Separation of both oxidized and phosphorylated VP proteoforms!



**HILIC method is patented!!**  
**US 2020/0131533 A1 (Apr 30, 2020)**

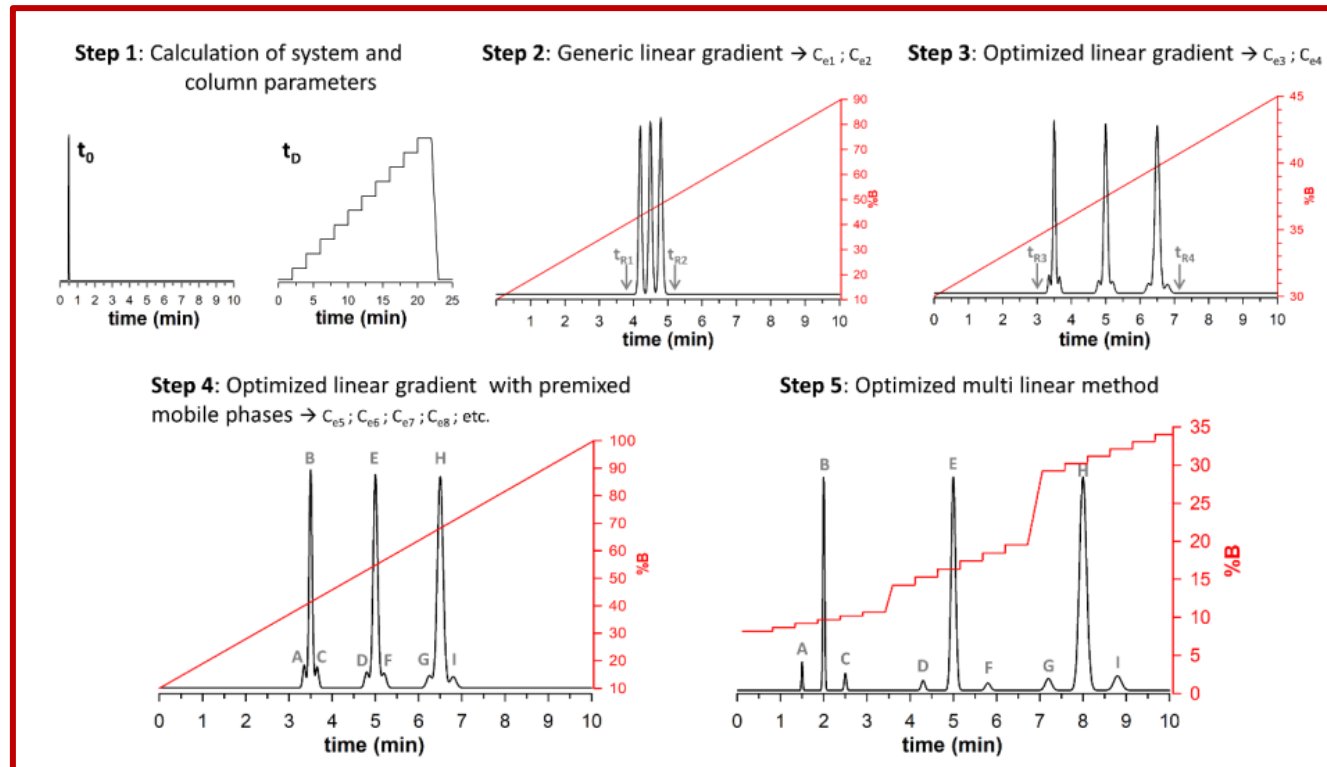
REF = Liu et al. JPBA 2020, 189, 113481

# Improved selectivity by using multi-isocratic elution



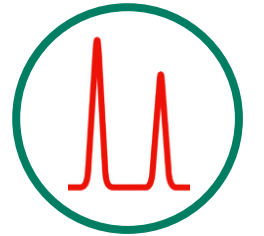
VPs are made of proteins,  
do they follow an “on/off” retention behaviour?

Use of the workflow in 5 steps (Murisier et al. Separations 2022, 9, 243)  
to develop multi-isocratic elution methods in RPLC, HILIC, and HIC.



Murisier et al. Separations 2022, 9, 243 & Aebischer et al. IJMS 2023, 24, 8503

# Improved selectivity by using multi-isocratic elution



VPs are made of proteins,  
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Use of the workflow in 5 steps (Murisier et al. Separations 2022, 9, 243)  
to develop multi-isocratic elution methods in RPLC, HILIC, and HIC.

## Patented HILIC mode (Liu et al. JPBA 2020)

Waters Glycoprotein BEH Amide column  
(300A, 1.7  $\mu\text{m}$ , 2.1 x 150mm);  
Injection volume = 3 x 1  $\mu\text{L}$  (multiple loading)  
MPA = 0.1%TFA in water  
MPB = 0.1%TFA in ACN  
T = 60 °C  
FR = 0.2 mL/min  
Gradient = linear with initial ramp at 85%B

## Optimized HILIC mode (Aebischer et al. IJMS 2023)

Waters Glycoprotein BEH Amide column  
(300A, 1.7  $\mu\text{m}$ , 2.1 x **50mm**);  
**Injection volume = 0.3  $\mu\text{L}$**   
MPA = 0.1%TFA in water  
**MPB = 0.1%TFA in ACN:IPA 80:20**  
**T = 40°C**  
**FR = 0.4 mL/min**  
**Gradient: multi-step gradient with premixed MPs**

Liu et al. JPBA 2020, 189, 113481 & Aebischer et al. IJMS 2023, 24, 8503



# Method development in HILIC mode: optimized linear gradient



A = [ACN/IPA, 80/20] + 0.1% TFA / H<sub>2</sub>O + 0.1% TFA (79/21)  
 B = [ACN/IPA, 80/20] + 0.1% TFA / H<sub>2</sub>O + 0.1% TFA (67/33)



15 min



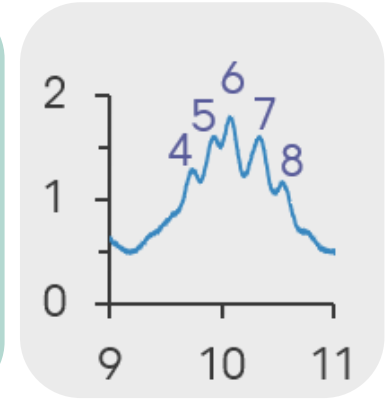
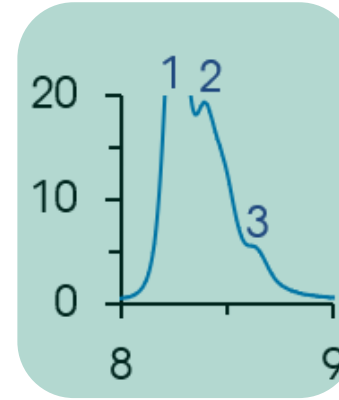
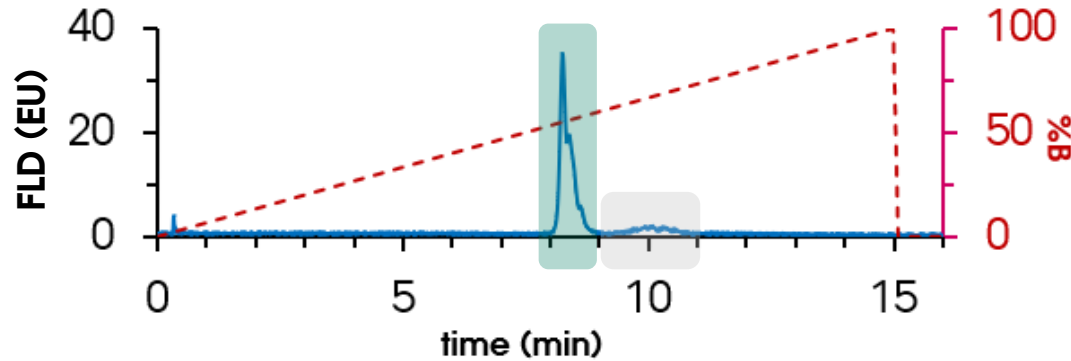
40 °C



0.4 mL/min

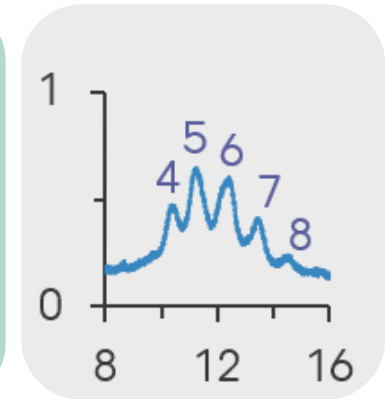
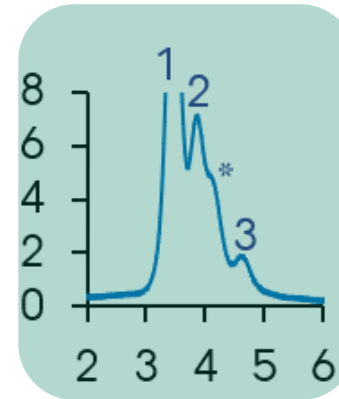
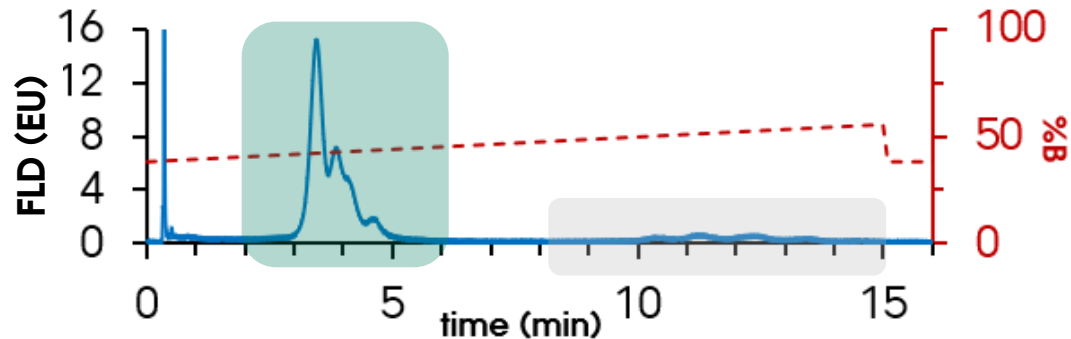
## 1. Generic linear gradient with premixed MPs

Time (min)	% B
0	0
15	100
15.1	0
20	0



## 2. Optimized linear gradient with premixed MPs

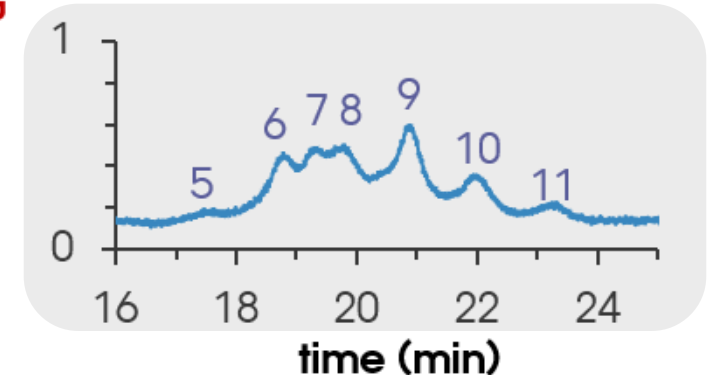
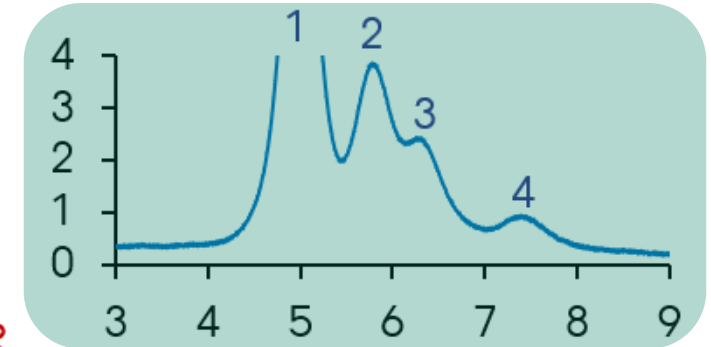
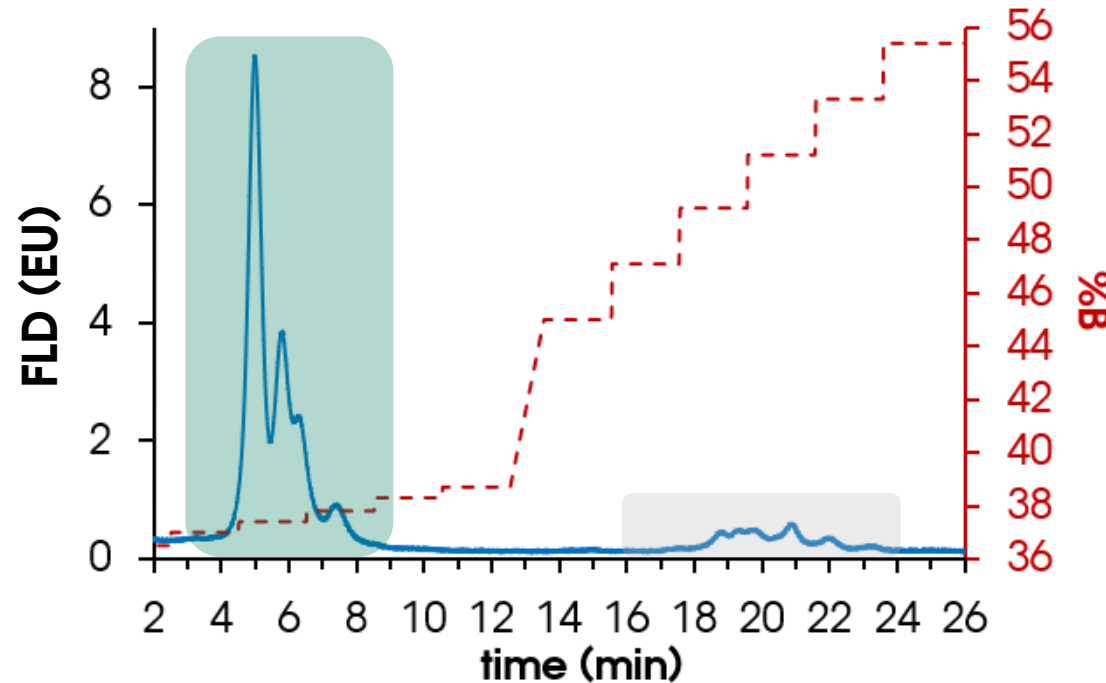
Time (min)	% B
0	38
15	56
15.1	38
20	38



Aebischer et al. JMS 2023, 24, 8503

## 3. Multi-step gradient with premixed MPs

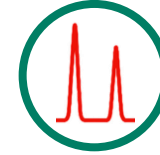
Time (min)	% B
0	36.5
0.50	36.5
2.50	36.5
2.51	37.0
4.51	37.0
4.52	37.4
6.52	37.4
6.53	37.8
8.53	37.8
8.54	38.3
10.54	38.3
10.55	38.7
12.55	38.7
13.55	45.0
15.55	45.0
15.56	47.1
17.56	47.1
17.57	49.2
19.57	49.2
19.58	51.2
21.58	51.2
21.59	53.3
23.59	53.3
23.60	55.4
26.10	55.4
26.20	36.5
40	36.5



**Separation of 11 viral protein variants at chromatographic level!**

# Conclusions

## Improved selectivity by using multi-isocratic elution



- Feasible for rAAV capsids
- Feasible for rAAV VPs
- Best selectivity gain when limited number of peaks to deal with (as for rAAV capsids application)
- Complex gradient program when dealing with large number of peaks (as for rAAV VPs application)

## ■ Acknowledgements



**UNIVERSITÉ  
DE GENÈVE**

**FACULTÉ DES SCIENCES**

Section des sciences  
pharmaceutiques

Dr Davy Guillaume

Megane Aebischer

Dr Thomas Bouvarel

Emmalyn Barrozo

Hugo Gizardin-Fredon



Dr Raphael Ruppert

Dr Markus Haindl

Carsten Elger

Dominik Kochardt

Waters

Dr Szabolcs Fekete



Centre de compétences  
**Chimie et Toxicologie  
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# THANK YOU FOR YOUR KIND ATTENTION!!



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